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Antidiabetic and Antihyperlipidaemic activity of *Polygala chinensis* L whole plant in alloxan induced diabetic rats

¹Alagammal M, ²Rajalakshmi K and ²Mohan VR*

¹Department of Botany, Government Siddha Medical College, Palayamkottai, Tamil Nadu.

²Ethnopharmacology Unit, Research Department of Botany, V. O. Chidambaram College, Tuticorin, Tamilnadu, India.

*Corresponding author: E-mail: vrmohanvoc@gmail.com

ABSTRACT

The ethanol extract of *Polygala chinensis* whole plant (Family: Polygalaceae) was investigated for its antidiabetic and antihyperlipidaemic effect in Wistar Albino rats. Diabetes was induced in Albino rats by administration of alloxan monohydrate (150mg/kg, i.p). The ethanol extracts of *Polygala chinensis* at a dose of 100 and 200mg/kg of body weight were administered at single dose per day to diabetes induced rats for a period of 14 days. The effect of ethanol extract of *Polygala chinensis* whole plant extract on blood glucose, serum insulin, urea, creatinine, glycosylated haemoglobin, serum lipid profile [total cholesterol (TR), triglycerides (TG), low density lipoprotein – cholesterol (LDL-C), very low density lipoprotein – cholesterol (VLDL-C), high density lipoprotein – cholesterol (HDL-C) and phospholipid (PL)] serum protein, albumin, globulin, serum enzymes [serum glutamate pyruvate transaminases (SGPT), and serum glutamate oxaloacetate transaminases (SGOT), and alkaline phosphatase (ALP)], were measured in the diabetic rats. The ethanol extract of *Polygala chinensis* whole plant elicited significant reductions of blood glucose (*P*<0.05), lipid parameters except HDL-C, serum enzymes and significantly increased HDL-C. The extracts also caused significant increase in serum insulin (*P*<0.05) in the diabetic rats. From the above results, it is concluded that ethanol extract of *Polygala chinensis* possesses significant antidiabetic and antihyperlipidaemic effects in alloxan induced diabetic rats.

Keywords: Antihyperlipidaemic, Antidiabetic, P. chinensis, Alloxan and Glibenclamide.

1. INTRODUCTION

According to World Health Organization (WHO) projections, the prevalence of diabetes is likely to increase by 35% by the year 2025 [1]. The prevalence of diabetes is estimated to reach 330 million by the year 2025, according to International Diabetes Federation, with the greatest potential increase being in Africa and Asia. This numerical increase will occur in developing countries. Thus by the year 2025 over 75% of people with diabetes will reside in developing countries, as compared to 62% in 1995^[2, 3]. Diabetes mellitus is a chronic metabolic disorder in the endocrine system with multiple characterized etiology, is by chronic hyperglycemia together with disturbances in carbohydrate, protein and fat metabolism results from a decrease in circulating concentration of insulin (insulin deficiency), a decrease in the response of peripheral tissues to insulin (insulin resistance) or both [4]. It is becoming the third "killer" mankind of after cancer and cardiovascular diseases, because of its high prevalence, morbidity and mortality ^[5]. The chronic hyperglycemia of diabetes is associated

with long term damage, dysfunction and failure of various orgens ^[6].

Recently the treatment of diabetes mainly involves a sustained reduction in hyperglycemia by the use of biguanides, thiazolidinediones, sulfonylureas, d-phenylalanine and α -glucosidase inhibitors in addition to insulin. However, due to unwanted side effects in the efficacies of these compounds are debatable and there is a demand for new compounds for the treatment of diabetes ^[6]. Hence plants have been suggested as a rich, as yet unexplored source of potentially useful antidiabetic drugs. Many traditional plants treatment for diabetes are used throughout the world.

According to the World Health Organization (WHO) estimates that 80% of people in developing countries depend on traditional medicine for their health needs, and 85% of traditional medicine involves the use of plant extracts. In other words, about 4 billion people in the world rely on plants as source of drugs. Plant drugs and herbal formulations are frequently considered to be less toxic and free from side effects than synthetic one ^[8].The antihyperglycemic effects of these plants are for their ability to restore the function of pancreatic tissues by increasing insulin output or inhibit the intestinal absorption of glucose or to the facilitation of metabolites in insulin dependent processes.

This has led researchers to continue their search for the "miracle drug" for treatment of diabetes from plants. Polygala chinensis L. belongs to Polygalaceae family. It is commonly known as "Siriyanangai". Genus Polygala was traditionally used by Americans to treat snake bites ^[9] and as an expectorant to treat cough and bronchitis. Polygala is considered as a powerful tonic herb ^[10] that can help to develop the mind and aid to creative thinking. Taking into the consideration of the medicinal importance of Polygala, the present study was conducted to investigate the antidiabetic and antihyperlipidaemic activities of ethanol extract of whole plant of Polygala chinensis in alloxan induced diabetic rats.

2. MATERIALS AND METHODS

2.1. Plant Material

The whole plant of *Polygala chinensis* were freshly collected from the well grown healthy plants inhabiting the natural forests of Maruthamalai, Coimbatore district, Tamil Nadu. The plant were identified and authenticated in Botanical Survey of India, Southern Circle, Coimbatore, Tamilnadu, India. A voucher specimen was deposited in Ethnopharmacology Unit, Research Department of Botany, V.O.Chidambaram College, Tuticorin, Tamilnadu.

2.2. Preparation of plant extract for phytochemical screening and antidiabetic studies

The *P. chinensis* whole plant was shade dried at room temperature and the dried whole plant was powdered in a Wiley mill. Hundred grams of powdered *P. chinensis* whole plant was packed in a Soxhlet apparatus and extracted with ethanol. The extracts were subjected to qualitative test for the identification of various phytochemical constituents as per the standard procedures ^[11, 12]. The ethanol extracts were concentrated in a rotary evaporator. The concentrated ethanol extract were used for antidiabetic studies.

2.3. Animals

Normal healthy male Wistar Albino rats (180- 240g) were housed under standard environmental conditions at temperature ($25\pm2^{\circ}$ C) and light and dark (12: 12 h). Rats were fed with standard pellet diet (Goldmohur brand, MS

Hindustan lever Ltd., Mumbai, India) and water *ad libitum*.

2.4. Acute Toxicity Study

Acute oral toxicity study was performed as per OECD – 423 guidelines (acute toxic class method), albino rats (n=6) of either sex selected by random sampling were used for acute toxicity study [13]. The animals were kept fasting for overnight and provided only with water, after which the extracts were administered orally at 5mg/kg body weight by gastric intubations and observed for 14 days. If mortality was observed in two out of three animals, then the dose administered was assigned as toxic dose. If mortality was observed in one animal, then the same dose was repeated again to confirm the toxic dose. If mortality was not observed, the procedure was repeated for higher doses such as 50,100, and 2000 mg/kg body weight.

2.5. Induction of Diabetes in Experimental animal

Rats were induced diabetes by the administration of simple intraperitoneal dose of alloxan monohydrate (150 mg/kg) ^[14]. Two days after alloxan injection, rats screened for diabetes having glycosuria and hypoglycemia with blood glucose level of 200-260 mg/100 ml were taken for the study. All animals were allowed free access to water and pellet diet and maintained at room temperature in plastic cages.

2.6. Experimental Design

In the present investigation, a total of 30 rats (24 diabetic surviving rats and 6 normal rats) were taken and divided into five groups of 6 rats each.

Group I: Normal untreated rats

Group II: Diabetic control rats

Group III: Diabetic rats given ethanol extract of *P. chinensis* whole plant (100mg/kg body weight)

Group IV: Diabetic rats given ethanol extract of *P. chinensis* whole plant (200mg/kg body weight)

Group V: Diabetic rats given standard drug glibenclamide (600µg/kg body weight).

The animals were sacrificed at the end of experimental period of 14 days by decapitation. Blood was collected, sera separated by centrifugation at 3000g for 10 minutes.

2.6.1. Estimation of insulin, glucose, urea, creatinine and glycosylated haemoglobin

Serum glucose was measured by the Otoluidine method ^[15]. Insulin level was assayed by Enzyme Linked Immuno Sorbant Assay (ELISA) kit ^[16]. Urea estimation was carried out by the method of Varley ^[17]; serum creatinine was estimated by the method of Owen *et al* ^[18]. Glycosylated haemoglobin (HBA₁C) estimation was carried out by a modified colorimetric method of Karunanayake and Chandrasekharan ^[19].

2.6.2. Estimation of protein, albumin, globulin, SGPT, SGOT, ALP

Serum protein ^[20] and serum albumins was determined by quantitative colorimetrically method by using bromocresol green. The total protein minus the albumin gives the globulin, serum glutamate pyruvate transaminase (SGPT) and serum glutamate oxaloacetate transaminase (SGOT) was measured spectrophotometrically by utilizing the method of Reitman and Frankel ^[21]. Serum alkaline phosphatase (ALP) was measured by the method of King and Armstrong ^[22].

2.6.3. Estimation of lipids and lipoprotein

Serum total cholesterol (TC) ^[23], total triglycerides (TG) ^[24], low density lipoprotein cholesterol (LDL-C), very low density lipoprotein cholesterol (VLDL-C) ^[25], high density lipoprotein cholesterol (HDL-C) ^[26] and phospholipids ^[27] were analyzed.

2.7. STATISTICAL ANALYSIS

The data were analyzed using student's ttest statistical methods. For the statistical tests a *P* values of less than 0.01 and 0.05 was taken as significant.

3. RESULTS AND DISCUSSION

The phytochemical screening of ethanol extract of P. chinensis whole plant revealed the presence of alkaloid, catechin, coumanin, flavonoid, tannin, saponin, steroid, phenol, glycoside, terpenoid and xanthoprotein. Acute toxicity study revealed the non-toxic nature of the ethanol extract of P. chinensis whole plant. Table 1 shows the levels of blood glucose, serum insulin, urea, creatinine and alvcosvlated haemoglobin of normal, diabetic rats and drug treated rats. The alloxan induced rats elicited significant rise in blood glucose from 83.16 to 231.84mg/dl (P<0.01) and a significant decrease in serum insulin from 14.27 to 5.21 (P<0.01). On the contrary, diabetic rats treated with ethanol extract of P. chinensis whole plant exhibited decrease in blood glucose and increase in serum insulin significantly (P<0.05) at a dose of 200mg/kg body weight. It was observed that ethanol extract of P. chinensis reversed these effects in diabetic rats. Glibenclamide, reference standard produced a significant (P<0.01) reduction in blood glucose and increase insulin compared to diabetic control.

Pancreas is the primary organ involved in sensing the organism's dietary and energetic states via glucose concentration in the blood and in response to elevated blood glucose; insulin is secreated [28]. Alloxan is one of the usual substances used for the inducing diabetes mellitus apart from streptozotocin. Alloxan has a destructive effect on the beta cells of the pancreas ^[29, 30]. Alloxan causes a massive reduction in insulin release by the destruction of β -cells of the islets of langerhans, there by inducing hyperglycaemia [31]. Insulin deficiency leads to various metabolic alterations in the animals viz increased blood glucose. The possible mechanism by which ethanol extract of *P. chinensis* whole plant brings about its hyperglycemic action may be induction of pancreatic insulin secretion from β -cells of islets of langerhans or due to enhanced transport of blood glucose to peripheral tissue [32]. Earlier many plants have been studied for their hypoglycemic and insulin release stimulatory effects [33-36].

significant elevation in А serum constituents, urea and creatinine were observed in alloxan induced diabetic rats (Group II), when compared to control rats. The ethanol extract of P. chinensis whole plant was administered orally (100mg/kg body weight-Group III 200mg/kg body weight-Group IV) to rats for 14 days, reversed to urea and creatinine level to near normal. The administration of glibenclamide (Group V) also decreased the levels of urea and creatinine to some extent. Alloxan is taken as inductions of an abnormal glomerular fraction where a simple injection of cisplation at a dose of 5mg/kg body weight in rabbits caused a marked reduction in the glomerular filtration rates, which was accompanied by an increase in the creatinine level, indicating the induction of acute renal failure. It is confirmed that there is a significant increase in serum creatinine in albino rats 14 days after alloxan administration. The present results show that, the treatment with ethanol extract of P. chinensis whole plant was effective in preventing alloxan induced increase in serum creatinine level when compared in the control. Alloxan induced diabetic rats showed significant increase (P<0.01) haemoglobin glycosylated (HBA_1C) level compared with normal rats. The ethanol extract of P. chinensis whole plant treated rats showed a significant decrease (P<0.05) in the content of glycosylated haemoglobin. Glycosylated haemoglobin determinations are self monitoring of blood glucose therefore play on important complementary roles for the management of diabetes mellitus [37].

The levels of serum protein, albumin and globulin of control, alloxan induced diabetic rats

Table -1: Effect of ethanol extract of P.chinensis whole plant on the serum insulin, glucose, urea,	
creatinine and HBA1C level of normal, diabetic induced and drug treated adult albino rats.	

Groups	Glucose (mg/dl)	Insulin(Mlu/ml)	Urea(mg/dl)	Creatinine (mg/dl)	HB A1C(%)
Ι	83.16±2.11	14.27±0.74	18.16±0.94	0.69±0.03	3.91±0.11
	231.84±18.43**	5.21±0.24**	36.69±1.14**	0.93±0.04*	8.59±0.14**
	108.26 ±1.26 ª	10.36± 0.36ª	18.29± 0.32ª	0.86 ±0.06 ª	6.24 ±0.26 ª
IV	133.84±2.08 [*] ^a	11.96±0.26ª	14.54±0.26ª	0.81±0.03	4.99±0.24ª
V	94.66±1.84 aa	13.63±0.18 aa	16.24±0.39ª	0.77±0.06	4.07±0.14ª

Each Value is SEM ± 6 individual observations * P < 0.05; ** P < 0.01 Compared normal control vs -Diabetic rats: a -P < 0.05; a - P < 0.01 Compared -Diabetic rats vs. drug treated.

Table -2: Effect of ethanol extract of *P.chinensis* whole plant on the serum protein, albumin, globulin, SGOT, SGPT and ALP level of normal, diabetic induced and drug treated adult albino rats.

Parameter	Protein (g/dl)	Albumin (g/dl)	Globulin(g/dl)	SGPT (u/I)	SGOT (u/I)	ALP (u/I)
Group I	8.14±0.17	4.50±0.18	3.64±0.13	18.31±0.61	21.66±0.34	120.16±4.11
Group II	5.27±0.11*	2.32±0.13*	2.95±0.16*	29.11±0.36**	34.16±0.71*	191.20±3.92**
Group III	6.24 ±0.13	3.24 ±0.12	3.00 ±0.08	24.61± 0.78	26.11± 0.32	154.16± 2.21
Group IV	7.94±0.14	4.44±0.17	3.50±0.21	15.11±0.36	22.14±0.33	129.63±1.14
Group V	8.01±0.31	4.23±0.14	3.78±0.25	18.16±0.23	24.30±0.14	124.66±2.74

Each Value is SEM ± 6 individual observations * *P* < 0.05; ** *P*<0.01 Compared normal control vs -Diabetic rats.

Table -3. Effect of *P. chinensis* extract on serum lipid profile in the normal, diabetic and drug treated rats.

Groups	TC(mg/dl)	TG(mg/dl)	HDL – C(mg/dl)	LDL - (mg/dl)	VLDL -C(mg/dl)	PL(mg/dl)
Ι	103.11±2.56	92.66±1.87	21.11±1.32	63.47±2.13	18.53±1.15	159.76±2.67
П	214.19±1.84**	178.26±2.19**	06.29±2.36**	92.25±2.56*	35.65±1.45	258.62±3.46
111	110.3±41.06	128.3±61.02	18.26±1.36	66.56±1.36	21.36±1.14	189.32±2.31
IV	96.16±1.13ªª	73.21±1.13ªª	21.74±1.23ª	57.78±1.22ª	15.64±1.27	153.58±2.35
V	108.26±1.33ª	84.64±1.72ª	30.16±1.30	61.18±1.32ª	16.92±1.14	164.35±2.88
VI	116.84±1.63ª	91.55±1.14 ^a	25.34±1.11ª	73.19±1.59 ^a	18.31±1.08	171.98±2.57

Each individual observations * *P*< 0.05; ** *P*<0.01 Compared normal control vs - Diabetic rats: a -*P* < 0.05; aa - *P*<0.01 Compared -Diabetic rats vs. drug treated. and drug treated rats were presented in Table 2. A significant reduction in serum protein, albumin and globulin were observed in alloxan induced diabetic rats (Group II) when compared to control (Group I) and glibenclamide treated rats (Group V). On administration of ethanol extract of *P. chinensis* to the diabetic rats, the levels of protein, albumin and globulin were found to be restored in normal. These results were in accordance with the effects of *Wattakaka volubilis* ^[38], *Senna auriculata* ^[39] and *Pterocarpus marsupium* ^[34] and *Eugenia floccose* ^[35].

Table 2 summarized the effect of alloxan on the activity of the hepatic marker enzymes in serum. In the present study, the levels of SGPT and SGOT in alloxan induced diabetic rats were elevated. It may be due to leaking out of enzymes from the tissues and migrating into the circulation by the adverse effect of alloxan [40]. AST and ALT were used as markers to assess the extent of liver damage in streptozotocin induced diabetic rats ^[41]. In this study, the ethanol extract of *P. chinensis* regulated the activity of SGPT and SGOT in liver of rats intoxicated with alloxan. The effect of glibenclamide on the recovery of hepatic enzyme activity in serum was very similar to that of the earlier study [42]. The restorations of SGPT and SGOT to their respective normal levels after treatment with both glibenclamide and ethanol extract of P. chinensis, further strengthen the antidiabegenic effect of this extract. Moreover, SGPT and SGOT levels also act as indicators of liver function and restoration of normal levels of these parameters indicate normal functioning of liver. Since the alloxan can also affect the liver by free radical mechanism.

In addition to the assessment of SGPT and SGOT levels during diabetes the measurement of enzymatic activities of phosphatases such as acid phosphatase (ACP) and alkaline phosphatase (ALP) is of clinical and toxicological importance as changes in their activities are indicative of tissue damage by toxicants. In the present study, serum ALP increased in alloxan induced diabetic rats (Table 2). Elevated level of this enzyme in diabetes may be due to extensive damage to liver in the experimental animal by alloxan. Treatment with ethanol extract of P. chinensis in alloxan induced diabetic rats produces a decline in ALP level. The levels of serum lipid profile, total cholesterol (TC), triglycerides (TG), LDL-C, VLDL-C and HDL-C in control, diabetic induced and drug treated rats were presented in Table 3. Alloxan induced rats showed significant increase in serum lipid profiles except HDL-C when compared with normal rats. The glibenclamide (Group V) and ethanol extract

of P. chinensis (Group III and IV) treated rats showed a significant decrease in the content of lipid profiles when compared with diabetic induced rats. Similarly HDL-C level decreased in alloxan induced diabetic rats when compared to normal rats. On administration of ethanol extract of *P. chinensis* and glibenclamide to the diabetic rats, HDL-C level was found to be restored to normal. The level of serum lipid profiles are usually raised in diabetic rats in the present study and such elevation represents risk factor for coronary heart diseases ^[43]. The hypolipidaemic effect may be due to inhibition of fatty acid synthesis [44]. In normal metabolism insulin activates the enzyme lipoprotein lipase and hydrolyses triglycerides and the deficiency in insulin results in inactivation of these enzymes thereby causing hypertriglyceridemia. The significant reduction of serum lipid levels in diabetic rats after *P. chinensis* treatment may be directly attributed to improvements in insulin levels. Phospholipids are present in cell membrane and make up vast majority of the surface lipoprotein forming a lipid bilayer that acts as an interface with both polar plasma environment and non-polar lipoprotein of lipoprotein core [45]. Increased phospholipids levels in tissues were reported by Venkateswaran et al [46]; Pari and Satheesh [47] in streptozotocin diabetic rats. Administration of ethanol extract of P. chinensis whole plant and glibenclamide decreased the levels of phospholipids.

4. CONCLUSION

The present study has shown that the ethanol extract of P. chinensis whole plant have antidiabetic and antihyperlipidaemic effects. From this study, the ethanol extract of *P. chinensis* whole plant is beneficial in controlling the blood glucose level, improves the lipid metabolism. The possible antidiabetic activity of the extracts might be due to stimulation of residual pancreatic insulin or by increasing peripheral utilization of glucose. Glycosides, flavonoids, tannins and alkaloids are active ingredients of hypoglycemic plant [48]. Flavonoids are reported to regenerate the damaged pancreatic beta cells [49]. Phenols have found to be effective antihyperglycemic agents ^[50]. In the present study, the phytochemical analysis of ethanol extract of P. chinensis clearly pointed out the presence of above said active phytochemicals. Further identification and isolation of these constituents may be fruitful.

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- 5. REFERENCES
- Boyle JP, Honeycutt AA, Narayan KM, Hoerges TJ, Geisis LS, Chen H and Thompson TJ. Projection of diabetes burden through 2050; Impact of champing demography and disease prevalence in the US. Diabetes care, 2001; 24: 1936-1940.
- Hillary KR and William HH. Global Burden of diabetes 1995-2005: Prevalence, Numerical estimates and projections. Diabetes care, 1998; 21:1414-1431.
- Ereyin O, Ebory P, Eyong E and Awofisayo O and Agboke A. Effects of Telfairia occidentalis on oral glucose tolerance in rates. Afr. J. Pharm. Pharmac., 2010;4: 368-372.
- Ashraj Ali M, Sagar HA, Khatun MCS, Azad AK, Begum K and Wahed MII. Antihyperglycemic and analgestic activities of ethanolic extract of *Carria fistula* (L.) Stem bark. Int. J. Pharmaceu. Sci. Res., 2012; 3: 416-423.
- Li WL, Zhengt HC, Bukuru J and De Kimpe N. Natural medicines used in the traditional Chinese medical system for therapy of diabetes mellitus. J. Ethnopharmacol., 2004; 92: 1-21.
- Lyra R, Oliveira M, Lins D, Cava L and Canti N. Prevention of type 2 diabetes mellitus. Arq. Bras. Endocrinol Metabo., 2006; 50: 239-249.
- 7. Moller DE. New drug targets for type 2 diabetes and the metabolic syndrome. Nature, 2001; 414: 821-827.
- 8. Kannur DM, Hukkeri VI and Akki KS. Antidiabetic activity of *Caesalpinia bonducella* seed extracts in rats Fitoterapia, 2006; 77: 546-549.
- Mc Guffin M, Hobbsc, Upton R (eds). American Herbal products Associations Botanical safety Hand book. Boca Raton, 1997; FL: CRC press P. 89.
- 10. Teeguarden R. Radiant health: The Ancient wisdom of the Chinese Tonic Herbs, New York: Warner Books, 1998; 194-195.
- 11. Brinda P, Sasikala P and Purushothaman KK. Pharmacognostic studies on *Merugan kizhangu*. Bull. Med. Ethnobot Res., 1981; 3: 84-96.
- 12. Lala PK. Lab manuals of Pharmacognosy CSI Publishers and Distributers, Kolkata 1993.
- 13. OECD. Organisation for Economic cooperation and Development). OECD

guidelines for the testing of chemicals/Section 4: Health Effects Test No. 423; Acute oral Toxicity- Acute Toxic Class method. OECD. Paris 2002.

- 14. Nagappa AN, Thakurdesai PA, Venkat Rao N and Sing J. Antidiabetic activity of *Terminalia catappa* Linn. fruits. J. Ethnopharmacol., 2003; 88: 45-50.
- Sasaki T, Masty S and Sonae A. Effect of acetic acid concentration on the colour reaction in the Otoluidine boric acid method for blood glucose estimation. Rinshbo Kagaku., 1972; 1: 346-353.
- Anderson L, Dinesen B, Jorgonsen PN, Poulsen F and Roder ME. Enzyme immune assay for intact human insulin in serum or plasma. Clin. Chem., 1993; 39: 578-582.
- Varley H. Practical clinical biochemistry, Arnold Heinemann Publication Pvt. Ltd., 1976; 452.
- Owen JA, Iggo JB, Scangrett FJ and Steward IP. Determination of creatinine in plasma serum, a critical examination. J. Biochem., 1954; 58: 426-437.
- 19. Karunanayake EH and Chandrasekharan NV. An evaluation of a colorimetric procedure for the estimation of glycosylated haemoglobin and establishment of reference values for Sri Lanka. J. Nat. Sci. Coun. Sri Lanka., 1985; 13: 235-258.
- 20. Lowry OH, Rosenbrough NJ, Farr AL and Randall RJ. Protein measurement with the folin's phenol reagent. J. Bio. Chem., 1951; 193: 265-275.
- 21. Reitman S and Frankel SA. Colorimetric method for the determination of serum glutamic oxaloacetic and glutamic pyruvic transaminases. Amer. J. Clin. Path., 1957; 28: 56-63.
- 22. King EJ and Armstrong AR. Determination of serum and bile phosphatase activity. Cannad. Med. Assoc. J., 1934; 31: 56-63.
- 23. Parekh AC and Jung. Cholesterol determination with ferric acetate, uranium acetate and sulphuric acid, ferrous sulphate reagent. Anal. Chem., 1970; 112: 1423-1427.
- 24. Rice EW. Triglycerides in Serum In: Standard Methods. Clinical Chemistry. 9ed Roderick MP, Academic press, New York. 1970; 215-222.
- 25. Friedwald WT, Levy RI and Fredrickson DS. Estimation of the concentration of low density lipoprotein cholesterol in plasma, without use

of the preparative ultra centrifuge. Clin. Chem., 1972; 18: 499-502.

- 26. Warnick GR, Nguyan T and Albers AA. Comparison of improved precipitation methods for quantification of high density lipoprotein cholesterol. Clin Chem., 1985; 31: 217.
- 27. Takayama M, Itoh S, Nagasaki T and Tanimizu I. A new enzymatic method for determination of serum phospholipids. Clin. Chem. Acta., 1977; 79: 93 – 98.
- 28. Edem DO. Hypoglycemic effects of ethanolic extracts of Alligator pear seed (*Persea Americana* Mill) in rats. European Journal of Scientific Research, 2009; 33: 669-678.
- 29. Prince SM, Menon VP. Hypoglycemic and other related action of *Tinospora cardifolia* roots in alloxan induced diabetic rats. J. Ethnopharmacol., 2000; 70: 9-15.
- Jelodar G, Mohsen M, Shahram S. Effect of walnut leaf, coriander and pomegranate on blood glucose and histopathology of pancreas of alloxan induced diabetic rats. African J Traditional complementary and Alternative Medicines, 2003; 3: 299-305.
- 31. Grover JK, Vats V and Rathi SS. Antihyperglycemic effect of Eugenia jambolana and Tinospora cardifolia in experimental diabetes and their effects on key metabolic enzymes involved in carbohydrate metabolism. Ethnopharmacol., 2000 73: 461-470.
- 32. Hakkim FL, Girija S, Senthilkumar R and Jalaludeen MD. Effect of aqueous and ethanol extracts of *Cassia auriculata* L. flowers on diabetic using alloxan induced diabetic rats. Int. J. Diabet. Matebol., 2007; 15: 100-106.
- 33. Pari L and Latha M. Effect of *Cassia auriculata flowers* on blood sugar levels, serum and tissue lipids in Streptozotocin Diabetic Rats. Sing. Med. J., 2002; 43: 617-621.
- 34. Maruthupandian A and Mohan VR. Antidiabetic, antihyperlipidaemic and antioxidant activity of *Pterocarpus marsupian* Roxb. in alloxan induced diabetic rats. Int. J. Pharm. Tech. Res., 2011; 3: 1681-1687.
- 35. Kala SMJ, Tresina PS and Mohan VR. Antioxidant, antihyperlipidaemic and antidiabetic activity of *Eugenia floccosa* Bedd leaves in Alloxan induced diabetic rats. J. Basic. Clin. Pharmacy, 2012; 3: 235-240.
- 36. Shajeela PS, Kalpanadevi V and Mohan VR. Potential antidiabetic, hypolipidaemic and

antioxidant effects of *Nymphaea pubercens* extract in Alloxan induced diabetic rats. J. Appl. Pharmacu. Sci., 2012; 2: 83-88.

- 37. Thai AC, Yeo PPB and Chan L. Glycosylated haemoglobin and diabetic control. Singapore Medicinal Journal, 1983; 24: 210-212.
- Maruthupandian A, Mohan VR, and Sampathraj R. Antidiabetic, antihyperlipidaemic and antioxidant activity of *Wattakaka volubilis* (L.f) Stapf leaves in alloxan induced diabetic rats. Int. J. Pharmaceut. Sci. Res., 2010; 1: 83-90.
- Shanmugasundaram R, Kalpana Devi V, Tresina Soris P, Maruthupandian A and Mohan VR. Antidiabetic, antihyperlipidaemic and antioxidant activity of *Senna auriculata* (L.) Roxb leaves in alloxan induced diabetic rats. Int. J. PharmTech. Res., 2011; 3: 747-756.
- Stanely P, Prince M and Menon V. Hypoglycemic and other related actions of *Tinospora cordifolia* roots in alloxan induced diabetic rats. J. Ethnopharmacol., 1999; 70: 9-15.
- Hwang HJ, Kim SW, Lim JM, Joo JH, Kim HO, Kim HM and Yun JW. Hypoglycemic effect of crude epoxypolysaccharides produced by a medicinal mushroom *Phellinus baumii* in streptozotocin induced diabetic rats. Life Science, 2005; 76: 3069-3080.
- 42. Preethi KC and Kuttan R. Hepato and reno protective action of *Calendula officinalis* L. flower extract. Ind. J. Exp. Biol., 2009; 47: 163-168.
- Mironova MA, Klein RL, Virella GL and Lopes-Virella MF. Anti-modified LDL antibodies, LD containing immune complexes and susceptibility of LDL to *in vitro* oxidation in patients with type 2 diabetes. Diabet., 2000; 49: 1033-1049.
- 44. Chi MS and Koh ET. Effect of garlic on lipid metabolism of rats fed with cholesterol or lard. J. Nutr., 1982; 112: 241-248.
- 45. Cohn RM and Roth KS. Lipid and lipoprotein metabolism In: Biochemistry and Diseases, Williams and Willkins publishers, Baltimore, 1996; p 280.
- Venkateswaran S, Pari L and Saravanan G. Effect of *Phaseolus vulgaris* on circulatory antioxidants and lipids in streptozotocininduced diabetic rats. J. Med. Food., 2002; 5: 97 – 104.

- 47. Pari L and Satheesh AM. Antidiabetic effect of *Boerhavia diffusa*: effect on serum and tissue lipids in experimental diabetes. J. Med. Food., 2004; 7: 472 476.
- Oliver B. Oral hypoglycaemic plants in West Africa. J. Ethnopharmacol., 1980; 2: 119-127.
- 49. Chakravarthy BK, Gupta S, Gambir SS and Gode KD. Pancreatic beta cell regeneration. A

novel antidiabetic mechanism of *Pterocarpus marsupium* Roxb. Ind. J. Pharmacol., 1980; 12: 123 – 127.

50. Manickam M, Ramanathan M, Farboodinary Jahromi MA, Chansouria JPN and Ray AB. Antihyperglycemic activity of phenolic form *Pterocarpus marsupium.* J. Nat. Prod., 1997; 60: 609-610.