

Synthesis of imidazole-schiff base analogues: SAR studies of potent antiglycation and urease activities

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ABSTRACT

A series of imidazole-Schiff base derivatives (**4-23**) were synthesized, spectroscopically characterized and evaluated for their *in vitro* antiglycation and urease inhibitory activities. The results shown that compounds **9, 10, 11, 15, 16, 22** and **23** showed excellent antiglycation and urease activities with IC₅₀ values are lower than the standards rutin and thiourea respectively. Preliminary structure-activity relationship revealed that the compounds **9, 10, 11, 15, 16, 22** and **23** with electron donating moiety (OH, OCH₃) were found to be excellent antiglycation and urease activity and compounds **5, 6, 7, 8, 12, 13, 14** and **21** with electron withdrawing groups (Cl, F, NO₂ and Br) were found to be least antiglycation and urease activity.

Keywords: Imidazole; Schiff's bases; Antiglycation; Urease activity; Electronic effect.

1. INTRODUCTION

Antiglycation agents play an important role in the treatment of diabetic complications. The number of effective antiglycation agents is still very limited. [1] The formation of advanced glycation end products (AGEs), is believed to play important roles in pathogenesis of diabetic and aging, [2] are considered these advanced glycation end products as vital mediators of approximately all diabetic complications. [3] Non-enzymatic reaction between reducing sugar and free amino group of proteins, leads to glycation product termed Amadori product. Schiff bases of glucose are produced primarily by the reaction between glucose and protein without any enzyme, this intermediate rearranges to Amadori products. [4] Few molecules have been found that can cleave AGEs cross-links and reverse the balanced process of diabetic complications. [5] S-Allylcysteine is an important component of aged garlic extract that inhibits the AGEs formation. [6] Many efforts have been made to develop new safe and sound synthetic antiglycation agents. [7]

The metalloenzyme urease (urea amidohydrolase; EC 3.5.1.5) catalyzes the hydrolysis urea into ammonia and carbon dioxide. It is present in a variety of plants, algae, fungi, and

bacteria. [8] Urease is involved in the pathogenesis of hepatic encephalopathy, hepatic coma urolithiasis, pyelo nephritis, ammonia, and urinary catheter encrustation. [8,9] It is also a major cause of pathologies induced by Helicobacter pylori as this allows bacteria to survive at the low pH of the stomach and hence plays an important role in producing peptic and gastric ulcers. [8] As a result, ureases have been identified as important targets in research both for human and animal health, as well as in agriculture.

Heterocyclic nucleus imparts an important function in medicinal chemistry and serves as a key template for the development of various therapeutic agents. [10] Imidazoles have occupied a unique position in heterocyclic chemistry, and its derivatives have attracted considerable interests in recent years for their versatile properties in chemistry and pharmacology. It improves pharmacokinetic characteristics of lead molecules and thus is used as a remedy to optimize solubility and bioavailability parameters of proposed poorly soluble lead molecules. The imidazole derivatives possess extensive spectrum of biological activities such as anti-inflammatory, [11] antioxidant, [12] antibacterial, [13] anticancer, [14] antitubercular [15]

and anti-HIV [16] activities. It is also present in the structure of many natural or synthetic drug molecules, that is, cimetidine, azomycin, and metronidazole. Imidazole-containing drugs have a broader scope in remedying various dispositions in clinical medicine.

Based on the above facts and in continuation of our drug development program, the present work involves the synthesis of a series of small and simple imidazole derived Schiff's base analogues as potential urease and antiglycation agents.

2. EXPERIMENTAL

2.1 Chemistry

General

All chemicals and reagents obtained from Sigma Aldrich (India), Merck (India) and Avra Synthesis (India) were used without further purification. Melting points were determined on a Superfit melting point apparatus (India) and are uncorrected. FT-IR was performed using a Jasco spectrometer (Japan) using nujol media. ¹H NMR (400 MHz) and ¹³C NMR (100 MHz) spectra were recorded on a Agilent Technologies (USA) using DMSO (*d*₆) as solvent. High resolution mass spectroscopic analysis was performed on a Bruker MicroTOF QII mass spectrometer in positive mode. Progress of the reaction was monitored by TLC using silica gel coated on glass plates with the solvent system comprising chloroform/methanol/acetic acid in the ratio 98:02:03 and the compounds on the TLC plates were detected by under UV light.

2.1.1. Ethyl 4-(2-hydroxypropan-2-yl)-2-propyl-1H-imidazole-5-carboxylate (2)

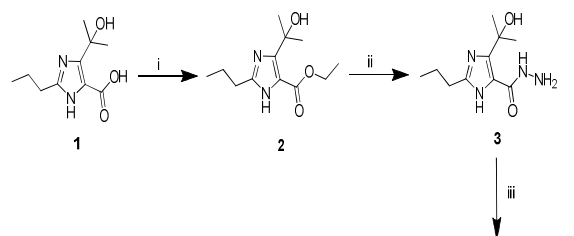
To a solution of imidazole (0.05 mol, 10.6 g) in ethanol (100 mL), trimethylsilylchloride (0.05 mol, 5.43 g) was added slowly. The reaction mixture was stirred for 4 hrs to complete the reaction (monitored by TLC). The solvent was removed under reduced pressure and the resultant precipitate was washed with ice cold water and filtered to yield the desired products **2**. Yield 10.8 g, 90.1%

Yield 10.8 g, 90.1 %, $R_f^a = 0.66$, $R_f^b = 0.71$, m.p. 184-185 °C, IR KBr (cm⁻¹): 1660, 3214, 3510; ¹H NMR (DMSO-*d*₆) δ ppm: 0.91-0.95 (t, 3H, CH₃), 1.30 (t, 3H, CH₃), 1.59 (s, 6H, 2CH₃), 1.68-1.73 (m, 2H, CH₂), 2.62-2.66 (t, 2H, CH₂), 4.30-4.35 (t, 2H, CH₂), 5.84 (s, 1H, OH), 9.9 (s, 1H, ring NH); HRMS *m/z*, (M+1): 241.1750

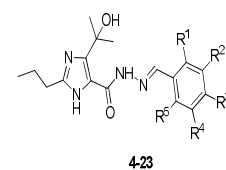
2.1.2. 4-(2-hydroxypropan-2-yl)-2-propyl-1H-imidazole-5-carbohydrazide (3)

To a solution of **2** (0.04 mol, 9.6 g) in ethanol (100 mL), hydrazine hydrate (0.048 mol,

2.4 g) was added. The reaction mixture was refluxed for 16 hrs for completion of the reaction (monitored by TLC). The solvent was removed under reduced pressure and cooled by adding ice cold water. The resulting precipitate was filtered, washed with cold water and recrystallized from ethanol to get the desired compounds **3**. Yield 7.7 g, 85.5 %



Reagents and Conditions:
 i) TMS-Cl, EtOH, rt, 4hr
 ii) NH₂-NH₂·H₂O, EtOH, reflux, 16 hr
 iii) Aromatic aldehydes, EtOH, acetic acid, EtOH, reflux, 8 hr



Entry	R ¹	R ²	R ³	R ⁴	R ⁵	Entry	R ¹	R ²	R ³	R ⁴	R ⁵
4	H	H	H	H	H	14	NO ₂	H	NO ₂	H	H
5	H	H	Cl	H	H	15	H	OH	OH	H	H
6	H	H	NO ₂	H	H	16	H	OCH ₃	OCH ₃	H	H
7	H	H	F	H	H	17	H	Br	OH	Br	H
8	H	H	Br	H	H	18	H	Br	OH	H	H
9	H	H	OCH ₃	H	H	19	H	Br	OCH ₃	H	H
10	H	H	OH	H	H	20	H	OCH ₃	OH	Br	H
11	H	OCH ₃	OH	H	H	21	Cl	H	H	H	F
12	Cl	H	Cl	H	H	22	H	OCH ₃	OCH ₃	OCH ₃	H
13	F	H	F	H	H	23	H	OH	OH	OH	H

Scheme - 1: Synthesis of target compounds 4-23.

Yield 7.7 g, 85.5 %, $R_f^a = 0.36$, $R_f^b = 0.40$, m.p. 201-202 °C, IR KBr (cm⁻¹): 1674, 2983, 3117, 3525; ¹H NMR (DMSO-*d*₆) δ ppm: 0.81-0.99 (t, 3H, CH₃), 1.22-1.58 (s, 6H, 2CH₃), 1.67-1.95 (m, 2H, CH₂), 2.58-2.62 (t, 2H, CH₂), 4.09-4.11 (d, 2H, NH₂), 7.09 (s, 1H, OH), 8.37 (t, 1H, NH), 8.84 (s, 1H, ring NH); HRMS *m/z*, (M+1): 227.1758

2.1.3. General procedure for the synthesis of Schiff's bases (4-23)

An equimolar amount of **3** (1 mmol) was dissolved in ethanol (10 mL/g of compound) and treated with appropriate aldehydes (1 mmol) in the presence of catalytic amount of glacial acetic acid. The reaction mixtures were refluxed for 7-8

hr and the completion of reaction was monitored by TLC. After completion of the reaction, the solvent was removed under reduced pressure and cooled by adding ice cold water. The resulting precipitate was filtered, washed with water and recrystallized from ethanol to obtain the desired Schiff's bases (4-23).

2.1.3.1. *N'*-Benzylidene-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (4)

Yield 88.40%, $R_f^a = 0.64$, $R_f^b = 0.71$, m.p. 189-190 °C, IR KBr (cm⁻¹): 1614, 1750, 3214, 3315, 3510; ¹H NMR (DMSO-*d*₆) δ ppm: 0.98 (s, 3H, CH₃), 1.27 (s, 6H, (CH₃)₂), 1.71 (m, 2H, CH₂), 2.92 (t, 2H, CH₂), 7.41-7.85 (m, 5H, Ar-H), 7.91 (s, 1H, -N=CH), 9.12 (s, 1H, OH), 10.21 (s, 1H, NH), 11.22 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.9, 24.1, 29.8, 31.3, 76.2, 128.6, 129.4, 131.1, 133.7, 136.4, 142.8, 143.6, 157.1, 160.1; HRMS m/z: 315.1548 [M+1]

2.1.3.2. *N'*-(4-Chlorobenzylidene-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (5)

Yield 91.10%, $R_f^a = 0.52$, $R_f^b = 0.56$, m.p. 159-160 °C, IR KBr (cm⁻¹): 1619, 1768, 3215, 3320, 3560; ¹H NMR (DMSO-*d*₆) δ ppm: 0.90 (s, 3H, CH₃), 1.29 (s, 6H, (CH₃)₂), 1.75 (m, 2H, CH₂), 2.87 (t, 2H, CH₂), 7.32-7.80 (m, 4H, Ar-H), 7.88 (s, 1H, -N=CH), 9.01 (s, 1H, OH), 10.21 (s, 1H, NH), 11.52 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.5, 24.4, 29.5, 31.8, 75.9, 128.9, 129.8, 132.1, 136.0, 136.9, 143.1, 144.2, 157.2, 160.1; HRMS m/z: 349.1236 [M+1], 351.6245 [M+3].

2.1.3.3. *N'*-(4-Nitrobenzylidene-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (6)

Yield 86.23%, $R_f^a = 0.42$, $R_f^b = 0.50$, m.p. 172-174 °C, IR KBr (cm⁻¹): 1606, 1730, 3212, 3355, 3588; ¹H NMR (DMSO-*d*₆) δ ppm: 0.85 (s, 3H, CH₃), 1.30 (s, 6H, (CH₃)₂), 1.74 (m, 2H, CH₂), 2.72 (t, 2H, CH₂), 7.60-8.12 (m, 4H, Ar-H), 7.88 (s, 1H, -N=CH), 8.90 (s, 1H, OH), 10.09 (s, 1H, NH), 11.12 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.6, 23.9, 28.6, 31.4, 76.2, 124.6, 125.7, 130.1, 136.1, 143.0, 143.9, 151.3, 157.8, 159.9; HRMS m/z: 360.6215 [M+1].

2.1.3.4. *N'*-(4-Fluorobenzylidene-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (7)

Yield 88.10%, $R_f^a = 0.49$, $R_f^b = 0.51$, m.p. 168-169 °C, IR KBr (cm⁻¹): 1610, 1770, 3250, 3370, 3568; ¹H NMR (DMSO-*d*₆) δ ppm: 0.88 (s, 3H, CH₃), 1.27 (s, 6H, (CH₃)₂), 1.77 (m, 2H, CH₂), 2.80 (t, 2H, CH₂), 7.12-7.42 (m, 4H, Ar-H), 7.95 (s, 1H, -N=CH), 8.51 (s, 1H, OH), 10.17 (s, 1H, NH), 11.30 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm:

12.9, 23.8, 29.1, 31.6, 75.8, 115.6, 128.7, 129.1, 136.7, 142.1, 144.5, 157.0, 157.9, 164.1; HRMS m/z: 333.4512 [M+1].

2.1.3.5. *N'*-(4-Bromobenzylidene-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (8)

Yield 86.20%, $R_f^a = 0.39$, $R_f^b = 0.42$, m.p. 185-187 °C, IR KBr (cm⁻¹): 1628, 1745, 3269, 3377, 3590; ¹H NMR (DMSO-*d*₆) δ ppm: 0.88 (s, 3H, CH₃), 1.32 (s, 6H, (CH₃)₂), 1.83 (m, 2H, CH₂), 2.69 (t, 2H, CH₂), 7.55-7.80 (m, 4H, Ar-H), 7.99 (s, 1H, -N=CH), 8.79 (s, 1H, OH), 10.56 (s, 1H, NH), 11.31 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.0, 24.1, 28.5, 30.5, 76.0, 124.9, 125.6, 131.1, 132.6, 136.5, 142.0, 143.5, 156.9, 160.1; HRMS m/z: 394.1254 [M+1], 396.5642 [M+3].

2.1.3.6. 4-(2-Hydroxypropan-2-yl)-*N'*-(4-methoxybenzylidene-2-propyl-1*H*-imidazole-5-carbohydrazide (9)

Yield 85.24%, $R_f^a = 0.45$, $R_f^b = 0.51$, m.p. 180-182 °C, IR KBr (cm⁻¹): 1606, 1733, 3310, 3395, 3555; ¹H NMR (DMSO-*d*₆) δ ppm: 0.92 (s, 3H, CH₃), 1.35 (s, 6H, (CH₃)₂), 1.90 (m, 2H, CH₂), 2.88 (t, 2H, CH₂), 3.78 (s, 3H, OMe), 7.10-7.82 (m, 4H, Ar-H), 7.91 (s, 1H, -N=CH), 8.56 (s, 1H, OH), 10.68 (s, 1H, NH), 11.12 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.5, 24.3, 29.4, 31.6, 55.6, 75.9, 114.9, 125.5, 130.6, 136.5, 142.8, 144.1, 157.0, 160.3, 163.5; HRMS m/z: 345.1254 [M+1].

2.1.3.7. *N'*-(4-Hydroxybenzylidene-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (10)

Yield 90.56%, $R_f^a = 0.35$, $R_f^b = 0.39$, m.p. 166-168 °C, IR KBr (cm⁻¹): 1610, 1740, 3260, 3380, 3565, 3590; ¹H NMR (DMSO-*d*₆) δ ppm: 0.90 (s, 3H, CH₃), 1.35 (s, 6H, (CH₃)₂), 1.90 (m, 2H, CH₂), 2.87 (t, 2H, CH₂), 6.80-7.45 (m, 4H, Ar-H), 7.88 (s, 1H, -N=CH), 8.90 (s, 1H, OH), 9.20 (s, 1H, OH), 10.61 (s, 1H, NH), 11.03 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.7, 24.6, 29.5, 31.3, 76.8, 116.9, 126.5, 130.2, 136.1, 142.5, 144.6, 157.5, 160.0, 160.9; HRMS m/z: 331.2364 [M+1].

2.3.1.8. *N'*-(4-Hydroxy-3-methoxybenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (11)

Yield 82.15%, $R_f^a = 0.41$, $R_f^b = 0.47$, m.p. 158-160 °C, IR KBr (cm⁻¹): 1612, 1780, 3336, 3341, 3562, 3585; ¹H NMR (DMSO-*d*₆) δ ppm: 0.88 (s, 3H, CH₃), 1.31 (s, 6H, (CH₃)₂), 1.86 (m, 2H, CH₂), 2.81 (t, 2H, CH₂), 3.78 (s, 3H, OMe), 6.91-7.62 (m, 3H, Ar-H), 8.02 (s, 1H, -N=CH), 9.12 (s, 1H, OH), 9.45 (s, 1H, OH), 10.37 (s, 1H, NH), 11.16 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 14.1, 25.2, 30.3, 31.0, 56.2, 76.1, 112.1, 117.0, 122.6, 130.6, 136.1, 143.5, 144.3, 149.8, 151.6, 157.6, 160.1; HRMS m/z: 361.4562 [M+1].

2.3.1.9. *N'*-(2,4-Dichlorobenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (12)

Yield 87.28%, $R_f^a = 0.43$, $R_f^b = 0.50$, m.p. 175-176 °C, IR KBr (cm⁻¹): 1622, 1758, 3312, 3379, 3545; ¹H NMR (DMSO-*d*₆) δ ppm: 0.89 (s, 3H, CH₃), 1.35 (s, 6H, (CH₃)₂), 1.80 (m, 2H, CH₂), 2.84 (t, 2H, CH₂), 7.20-7.82 (m, 3H, Ar-H), 8.10 (s, 1H, -N=CH), 9.52 (s, 1H, OH), 10.52 (s, 1H, NH), 11.13 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.3, 24.5, 30.6, 31.4, 76.3, 126.9, 128.2, 129.4, 129.9, 131.4, 132.0, 136.1, 140.5, 142.3, 157.6, 160.3; HRMS *m/z*: 384.1546 [M+1], 386.4569 [M+3].

2.3.1.10. *N'*-(2,4-Difluorobenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (13)

Yield 84.20%, $R_f^a = 0.46$, $R_f^b = 0.52$, m.p. 181-182 °C, IR KBr (cm⁻¹): 1630, 1755, 3324, 3318, 3569; ¹H NMR (DMSO-*d*₆) δ ppm: 0.85 (s, 3H, CH₃), 1.32 (s, 6H, (CH₃)₂), 1.83 (m, 2H, CH₂), 2.79 (t, 2H, CH₂), 6.92 (s, 1H, Ar-H), 7.13-7.70 (m, 2H, Ar-H), 7.84 (s, 1H, -N=CH), 8.99 (s, 1H, OH), 10.60 (s, 1H, NH), 11.17 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 14.3, 24.9, 29.5, 31.1, 76.1, 111.3, 112.9, 113.4, 132.1, 136.4, 142.8, 143.6, 157.2, 160.3, 161.4, 163.2; HRMS *m/z*: 351.4521 [M+1].

2.3.1.11. *N'*-(2,4-Dinitrobenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (14)

Yield 81.98%, $R_f^a = 0.51$, $R_f^b = 0.57$, m.p. 191-192 °C, IR KBr (cm⁻¹): 1622, 1768, 3375, 3398, 3514; ¹H NMR (DMSO-*d*₆) δ ppm: 0.89 (s, 3H, CH₃), 1.29 (s, 6H, (CH₃)₂), 1.77 (m, 2H, CH₂), 2.86 (t, 2H, CH₂), 7.92 (s, 1H, N=CH), 8.20-8.40 (m, 2H, Ar-H), 8.84 (s, 1H, Ar-H), 9.99 (s, 1H, OH), 10.29 (s, 1H, NH), 11.03 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.9, 23.9, 29.3, 31.5, 76.4, 120.5, 130.4, 131.2, 133.1, 136.1, 142.4, 143.9, 148.2, 151.6, 157.2, 160.3; HRMS *m/z*: 405.3542 [M+1].

2.3.1.12. *N'*-(3,4-Dihydroxybenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (15)

Yield 86.77%, $R_f^a = 0.32$, $R_f^b = 0.37$, m.p. 166-167 °C, IR KBr (cm⁻¹): 1610, 1710, 3318, 3374, 3520, 3599; ¹H NMR (DMSO-*d*₆) δ ppm: 0.84 (s, 3H, CH₃), 1.22 (s, 6H, (CH₃)₂), 1.82 (m, 2H, CH₂), 2.78 (t, 2H, CH₂), 6.80-7.45 (m, 3H, Ar-H), 7.88 (s, 1H, -N=CH), 8.84 (s, 1H, OH), 9.57 (s, 2H, OH), 10.56 (s, 1H, NH), 11.09 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.8, 23.7, 29.3, 31.0, 76.1, 116.2, 118.3, 123.5, 131.1, 136.3, 142.7, 143.1, 148.1, 151.0, 157.3, 160.7; HRMS *m/z*: 347.6524 [M+1].

2.3.1.13. *N'*-(3,4-Dimethoxybenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (16)

Yield 83.15%, $R_f^a = 0.44$, $R_f^b = 0.51$, m.p. 157-158 °C, IR KBr (cm⁻¹): 1612, 1770, 3317, 3384, 3566; ¹H NMR (DMSO-*d*₆) δ ppm: 0.89 (s, 3H, CH₃), 1.20 (s, 6H, (CH₃)₂), 1.90 (m, 2H, CH₂), 2.90 (t, 2H, CH₂), 3.82 (s, 6H, 2OMe), 6.92-7.51 (m, 3H, Ar-H), 7.93 (s, 1H, -N=CH), 8.99 (s, 1H, OH), 10.12 (s, 1H, NH), 11.16 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.9, 23.8, 29.4, 31.4, 56.4, 76.7, 109.3, 111.3, 122.6, 130.8, 136.8, 142.8, 144.2, 150.1, 152.4, 157.8, 160.4; HRMS *m/z*: 375.2654 [M+1].

2.3.1.14. *N'*-(3,5-Dibromo-4-hydroxybenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (17)

Yield 85.10%, $R_f^a = 0.40$, $R_f^b = 0.44$, m.p. 168-169 °C, IR KBr (cm⁻¹): 1608, 1778, 3330, 3398, 3547; ¹H NMR (DMSO-*d*₆) δ ppm: 0.93 (s, 3H, CH₃), 1.28 (s, 6H, (CH₃)₂), 1.94 (m, 2H, CH₂), 2.84 (t, 2H, CH₂), 7.60-7.72 (m, 2H, Ar-H), 7.88 (s, 1H, -N=CH), 8.78 (s, 1H, OH), 9.45 (s, 1H, OH), 10.19 (s, 1H, NH), 11.12 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.4, 23.6, 29.5, 31.3, 76.3, 110.3, 129.3, 130.6, 136.8, 142.8, 145.8, 157.2, 158.1, 160.6; HRMS *m/z*: 489.2314 [M+1], 491.2654 [M+3].

2.3.1.15. *N'*-(3-Bromo-4-hydroxybenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (18)

Yield 87.10%, $R_f^a = 0.46$, $R_f^b = 0.51$, m.p. 174-175 °C, IR KBr (cm⁻¹): 1616, 1788, 3320, 3399, 3565; ¹H NMR (DMSO-*d*₆) δ ppm: 0.89 (s, 3H, CH₃), 1.33 (s, 6H, (CH₃)₂), 1.87 (m, 2H, CH₂), 2.74 (t, 2H, CH₂), 6.90-7.77 (m, 3H, Ar-H), 7.97 (s, 1H, -N=CH), 8.88 (s, 1H, OH), 9.45 (s, 1H, OH), 10.82 (s, 1H, NH), 11.13 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 14.4, 24.1, 29.8, 31.6, 76.4, 113.3, 118.6, 128.4, 129.4, 130.4, 136.7, 142.7, 145.4, 157.4, 158.4, 160.9; HRMS *m/z*: 410.2654 [M+1], 412.2654 [M+3].

2.3.1.16. *N'*-(3-Bromo-4-methoxybenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (19)

Yield 88.17%, $R_f^a = 0.51$, $R_f^b = 0.57$, m.p. 179-181 °C, IR KBr (cm⁻¹): 1607, 1729, 3314, 3347, 3558; ¹H NMR (DMSO-*d*₆) δ ppm: 0.91 (s, 3H, CH₃), 1.37 (s, 6H, (CH₃)₂), 1.88 (m, 2H, CH₂), 2.82 (t, 2H, CH₂), 3.81 (s, 3H, OMe), 6.92-7.71 (m, 3H, Ar-H), 7.89 (s, 1H, -N=CH), 9.10 (s, 1H, OH), 10.22 (s, 1H, NH), 11.14 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ ppm: 13.5, 24.3, 29.7, 31.3, 56.4, 76.5, 111.3, 112.6, 128.1, 129.0, 129.4, 136.0, 142.3,

144.1, 157.1, 158.3, 160.6; HRMS m/z: 424.1264 [M+1], 426.4597 [M+3].

2.3.1.17. *N'*-(3-Bromo-4-hydroxy-5-methoxybenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (20)

Yield 89.27%, $R_f^a = 0.42$, $R_f^b = 0.47$, m.p. 165-168 °C, IR KBr (cm^{-1}): 1611, 1735, 3322, 3354, 3560; ^1H NMR ($\text{DMSO-}d_6$) δ ppm: 0.87 (s, 3H, CH_3), 1.34 (s, 6H, $(\text{CH}_3)_2$), 1.84 (m, 2H, CH_2), 2.77 (t, 2H, CH_2), 3.77 (s, 3H, OMe), 7.32-7.42 (m, 2H, Ar-H), 7.87 (s, 1H, $-\text{N}=\text{CH}$), 9.10 (s, 1H, OH), 9.88 (s, 1H, OH), 10.21 (s, 1H, NH), 11.22 (s, 1H, NH); ^{13}C NMR ($\text{DMSO-}d_6$) δ ppm: 13.2, 24.1 29.6, 31.8, 56.2, 76.1, 111.0, 114.4, 122.1, 129.9, 136.7, 142.1, 143.4, 145.6, 153.6, 157.4, 159.9; HRMS m/z: 440.1654 [M+1], 442.1564 [M+3].

2.3.1.18. *N'*-(2-Chloro-6-fluorobenzylidene)-4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carbohydrazide (21)

Yield 86.38%, $R_f^a = 0.52$, $R_f^b = 0.59$, m.p. 170-171 °C, IR KBr (cm^{-1}): 1602, 1758, 3310, 3359, 3566; ^1H NMR ($\text{DMSO-}d_6$) δ ppm: 0.89 (s, 3H, CH_3), 1.39 (s, 6H, $(\text{CH}_3)_2$), 1.89 (m, 2H, CH_2), 2.90 (t, 2H, CH_2), 7.22-7.49 (m, 3H, Ar-H), 7.90 (s, 1H, $-\text{N}=\text{CH}$), 9.28 (s, 1H, OH), 10.12 (s, 1H, NH), 11.01 (s, 1H, NH); ^{13}C NMR ($\text{DMSO-}d_6$) δ ppm: 13.8, 24.3 29.1, 31.0, 76.0, 113.8 118.4, 125.7, 134.5, 135.4, 136.9, 142.0, 143.7, 156.5, 160.5, 161.5; HRMS m/z: 367.4521 [M+1], 369.2451 [M+3].

2.3.1.19. 4-(2-Hydroxypropan-2-yl)-2-propyl-*N'*-(3,4,5-trimethoxybenzylidene)-1*H*-imidazole-5-carbohydrazide (22)

Yield 85.41%, $R_f^a = 0.42$, $R_f^b = 0.48$, m.p. 168-169 °C, IR KBr (cm^{-1}): 1610, 1766, 3352, 3369, 3588; ^1H NMR ($\text{DMSO-}d_6$) δ ppm: 0.88 (s, 3H, CH_3), 1.36 (s, 6H, $(\text{CH}_3)_2$), 1.84 (m, 2H, CH_2), 2.77 (t, 2H, CH_2), 3.82 (s, 9H, 3OMe), 7.12-7.18 (m, 2H, Ar-H), 7.99 (s, 1H, $-\text{N}=\text{CH}$), 9.38 (s, 1H, OH), 10.11 (s, 1H, NH), 11.56 (s, 1H, NH); ^{13}C NMR ($\text{DMSO-}d_6$) δ ppm: 13.2, 24.8 29.7, 31.6, 56.2, 60.6, 76.7, 104.2, 128.4, 136.4, 141.5, 142.4, 144.9, 153.5, 156.0, 160.4; HRMS m/z: 405.1265 [M+1].

2.3.1.20. 4-(2-Hydroxypropan-2-yl)-2-propyl-*N'*-(3,4,5-trihydroxybenzylidene)-1*H*-imidazole-5-carbohydrazide (23)

Yield 84.09%, $R_f^a = 0.30$, $R_f^b = 0.34$, m.p. 174-175 °C, IR KBr (cm^{-1}): 1615, 1719, 3349, 3359, 3562; ^1H NMR ($\text{DMSO-}d_6$) δ ppm: 0.84 (s, 3H, CH_3), 1.30 (s, 6H, $(\text{CH}_3)_2$), 1.80 (m, 2H, CH_2), 2.81 (t, 2H, CH_2), 5.01 (s, 1H, OH), 6.88-7.10 (m, 2H, Ar-H), 7.78 (s, 1H, $-\text{N}=\text{CH}$), 8.12 (s, 2H, 2OH), 9.38 (s, 1H, OH), 10.11 (s, 1H, NH), 11.56 (s, 1H, NH); ^{13}C NMR ($\text{DMSO-}d_6$) δ ppm: 13.1, 24.5 29.6,

31.0, 76.4, 108.2, 129.4, 136.1, 138.5, 141.7, 143.3, 146.1, 157.5, 160.1; HRMS m/z: 363.1265 [M+1]

2.4. Biology

2.4.1. *In vitro* Antiglycation assay [17]

Sodium phosphate buffer (pH 7.4) was prepared by mixing Na_2HPO_4 and NaH_2PO_4 (67 mM) containing sodium azide (3 mM); phosphate buffer saline (PBS) was prepared by mixing NaCl (137 mM) + Na_2HPO_4 (8.1 mM) + KCl (2.68 mM) + KH_2PO_4 (1.47 mM) and pH 10 was adjusted with NaOH (0.25 mM), while BSA (10 mg/mL) and anhydrous glucose (50 mg/mL) solutions were prepared in sodium phosphate buffer.

Bovine serum albumin (10 mg/mL) was incubated with glucose anhydrous (50 mg/mL) in sodium phosphate buffer (pH 7.4). DMSO used for dissolving the compounds was found to have no effect on the reaction at <2% (v/v). Glycated control contains 20 μL BSA + 20 μL glucose + 20 μL sodium phosphate buffer, while blank control contains 20 μL BSA and 40 μL sodium phosphate buffer. The mixture was incubated at 37°C for 7 days. After incubation, 6 μL (100%) of TCA was added into each well and centrifuged (15,000 rpm) for 4 min at 4 °C. After centrifugation, the pellets were rewashed with 60 μL (10%) of TCA. The supernatant containing glucose, inhibitor and interfering substance was removed and pellet containing advanced glycated end product-BSA were dissolved in 60 μL phosphate buffer solution (PBS). Evaluation of fluorescence spectrum (excitation 370 nm), and change in fluorescence intensity (excitation 370 nm to emission 440 nm), based on AGEs were monitored by using spectrofluorimeter (RF-1500, Shimadzu, Japan). % Inhibition was calculated using the formula:

$$\% \text{ Inhibition} = 1 - \frac{\text{Fluorescence of sample}}{\text{Fluorescence of glycated sample}} \times 100$$

Rutin was used as the standard antiglycation agent.

2.4.2. *In vitro* urease inhibition assay [18]

Reaction mixtures comprising 25 μL of jack bean urease enzyme (10mg/ml of 0.2M SPB) solution and 55 μL of buffers containing 100 μM urea were incubated with 5 μL of the test compounds (0.5-500 μM concentration) at 30°C for 15 min in 96-well plates. Urease activity was determined by measuring ammonia production using the indophenol method as described by Weatherburn [20]. Briefly, 45 μL each of phenol reagent (1% w/v phenol and 0.005% w/v sodium nitroprusside) and 70 μL of alkali reagent (0.5% w/v NaOH and 0.1% active chloride NaOCl) were added to each well. The increasing absorbance at 630 nm was measured after 50 min using a micro

plate reader (RF-1500, Shimadzu, Japan). All the reactions were performed in triplicate in a final volume of 200 μ l. The entire assays were performed at pH 6.8. Percentage inhibition was calculated from the formula

$$\% \text{ Inhibition} = 1 - \left(\frac{\text{OD test well}}{\text{OD control}} \right) \times 100$$

Thiourea was used as the standard inhibitor of urease assay.

3. RESULTS AND DISCUSSION

3.1. Chemistry

Syntheses of the desired compounds were achieved according to the steps illustrated in

3.2. Scheme

4-(2-hydroxypropan-2-yl)-2-propyl-1*H*-imidazole-5-carboxylic acid (**1**) were ethylated using trimethylsilylchloride (TMS-Cl) and ethanol at room temperature, which upon reaction with excess of hydrazine hydrate afforded the corresponding imidazole hydrazide (**3**). The Schiff's bases (**4-23**) were obtained by reacting **3** with different aromatic aldehydes in presence of catalytic amount of glacial acetic acid. All the derivatives were obtained in high yield and the methods employed are very simple. The structures of all the newly synthesized compounds including intermediates were confirmed by IR, ¹HNMR, ¹³CNMR and mass spectral analysis. The formations of ethyl ester (**2**) were confirmed by the appearance of a triplet at 1.30-1.33 δ for ethyl CH₃ and multiplet at 4.30-4.35 δ for ethyl CH₂ and absence of COOH proton peak at 11.80 δ in ¹HNMR spectrum. In IR spectra, bands at 2983 and 3117cm⁻¹ for NH₂-NH groups indicates the conversion of ethyl esters into hydrazides. The formation of Schiff's bases were confirmed by the presence of absorption at 1604-1635 for imines i.e., -N=CH- in IR spectra. The presence of all requisite peaks and absence of extraneous peaks in ¹HNMR and ¹³CNMR confirms the structures.

3.3. Biological activity

3.3.1. Antiglycation and Urease activity

We have synthesized imidazole-Schiff base (**4-23**) analogues and their subjected to antiglycation and urease inhibitory activities. The results obtained are presented in table 1 and the data represents average values from triplicate runs. Activity of the test compounds was compared with rutin and thiourea, which served as reference standards for antiglycation and urease inhibitory activities respectively. Most of the synthesized compounds showed potent antiglycation and urease inhibition activity.

Compounds **9, 10, 11, 15, 16, 22** and **23** showed excellent antiglycation and urease activities with IC₅₀ values are lower than the standards rutin and thiourea respectively. The remaining compounds **4-8, 12-14, 17, 18, 19-21** and **24** showed least antiglycation and urease inhibition activities with IC₅₀ values are more than standards. Results revealed that compounds containing electron donating (OH and OCH₃) groups (**9, 10, 11, 15, 16, 22** and **23**) are more active than electron withdrawing (Cl, NO₂, F and Br) groups (**5-8, 12-14, 17, 18, 19-21** and **24**).

Table - 1: Antiglycation and urease inhibition of synthesized compounds

Entry	Antiglycation activity ^a (IC ₅₀ μ M)	Urease inhibitory activity ^a (IC ₅₀ μ M)
1	176 \pm 1.2	44 \pm 0.3
2	Inactive	Inactive
3	Inactive	Inactive
4	158 \pm 1.2	40 \pm 0.4
5	90 \pm 0.6	34 \pm 1.2
6	111 \pm 1.0	30 \pm 0.4
7	55 \pm 0.1	24 \pm 0.6
8	120 \pm 1.8	29 \pm 1.0
9	32 \pm 0.9	18 \pm 1.6
10	26 \pm 1.5	15 \pm 0.9
11	19 \pm 0.5	12 \pm 0.3
12	126 \pm 0.7	31 \pm 0.4
13	74 \pm 1.6	36 \pm 1.0
14	141 \pm 0.3	39 \pm 1.6
15	17 \pm 1.3	11 \pm 0.6
16	21 \pm 0.8	15 \pm 0.1
17	51 \pm 0.2	21 \pm 1.3
18	45 \pm 1.3	25 \pm 0.8
19	55 \pm 0.9	28 \pm 0.3
20	39 \pm 0.4	20 \pm 0.6
21	60 \pm 0.4	30 \pm 1.3
22	15 \pm 0.1	8 \pm 0.5
23	10 \pm 0.2	6 \pm 0.4
Rutin	41 \pm 0.45	-
Thiourea	-	21 \pm 0.53

^a Values are mean of three determinations, the ranges of which are <5% of the mean in all cases.

The antiglycation and urease activities of Schiff bases of imidazoles C₆H₅CHO without any substituent on the phenyl ring were almost same

as that of imidazole moiety. The introduction of OH or OCH₃ on the phenyl ring drastically increases the antiglycation and urease activity and even the magnitude of increase is in accordance with the number of OH and OCH₃ groups. The Schiff bases with three OH groups or three OCH₃ groups or two OH or two OCH₃ groups on phenyl rings exhibited striking antiglycation and urease activities. The present investigation reveals that OH and OCH₃ were capable of increasing the activity even in the presence of electron withdrawing groups such as F, Cl, Br and NO₂. On the basis of present observation, it was very clear that the presence of OH or OCH₃ or both makes the molecules as potent antiglycation and urease activities. The compounds with electron withdrawing groups F, Cl, Br and NO₂ showed least antiglycation and urease activities. To study the structure activity relationship (SAR), compounds having OH (phenolic) and OCH₃ (anisole) groups in the phenyl ring (**9**, **10**, **11**, **15**, **16**, **22** and **23**) were found to be the most potent antiglycation and urease activities. In phenyl ring the number of hydroxy and methoxy group increases the activities also increases. The activity order is OCH₃ > OH > 2 OCH₃ > 2 OH > 3 OCH₃ > 3 OH. On the other hand, it seems to be interesting to point out that among the halogen substituted derivatives (Cl, F, NO₂ and Br), compounds containing fluoro proved to be an activity enhancer which is in well agreement with earlier reports. [19,20] This may be due to the more electron withdrawing nature of the fluoro compared to other halogens.

4. CONCLUSION

In conclusion, we have designed and synthesized a series of small and simple imidazole derived Schiff base analogues with different groups in benzene ring. From the biological activities studies, compounds **9**, **10**, **11**, **15**, **16**, **22** and **23** with OH and OCH₃ groups in benzene ring (electron donating) exhibited excellent antiglycation and urease inhibition activity. Compounds **5**, **6**, **7**, **8**, **12**, **13**, **14** and **21** with Cl, F, NO₂ and Br in benzene ring (electron withdrawing) showed least antiglycation and urease inhibition activity.

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5. REFERENCES

1. Ahmed N. Advanced glycation endproducts-role in pathology of diabetic complications. **Diabetes Research and Clinical Practice**. 2005; 67: 3-21.
2. Brownlee M. Glycation and diabetic complications. **Diabetes**. 1994; 43: 836-841.
3. Peppas M, Uribarri J and H. Vlassara H. Glucose advanced glycation end products and diabetes complications: What is new and what works. **Diabetes**. 2003; 21: 186-187.
4. Monnier VM. Enzymatic deglycation of proteins. **Archives of Biochemistry and Biophysics**. 2003; 419: 1-15.
5. Vasan S, Foiles P and Founds H. Therapeutic potential of breakers of advanced glycation end product-protein crosslinks. **Archives of Biochemistry and Biophysics**. 2003; 419: 89-96.
6. Hunt JV, Bottoms MA and Mitchinson MJ. Oxidative alterations in the experimental glycation model of diabetes mellitus are due to protein-glucose adduct oxidation. Some fundamental differences in proposed mechanisms of glucose oxidation and oxidant production. **Journal of Biochemistry**. 1993; 291: 529-535.
7. Singh R, Barden A, Mori T and Beilin L. Advanced glycation end-products: a review. **Diabetes**. 2001; 44: 129-146.
8. Krajewska B. Ureases: roles, properties and catalysis. **Wiad Chemistry**. 2002; 56: 223-253.
9. Mobley HLT, Island MD, and Hausinger RP. Molecular biology of microbial ureases. **Microbiological Reviews**. 1995; 59: 451-480.
10. Kashyap SJ, Sharma PK, Garg VK, Dudhe R and Kumar N. Review on synthesis and various biological potential of thiazolopyrimidine derivatives. **Journal of Advanced Scientific Research**. 2011; 2: 18-24.
11. Husain A, Drabu S, Kumar N, Alam MM and Bawa S. Synthesis and biological evaluation of di- and tri-substituted imidazoles as safer anti-inflammatory-antifungal agents. **Journal of Pharmacy and Bioallied Sciences**. 2013; 5: 154-161.
12. Sorrenti V, Salerno L, Giacomo C, Acquaviva R, Siracusa MA and A. Vanella A. Imidazole derivatives as antioxidants and selective inhibitors of Nnos. **Nitric Oxide**. 2006; 14: 45-50.
13. Vijesh AM, Isloor AM, Telkar S, Peethambar SK, Rai S and Isloor N. Synthesis, characterization and antimicrobial studies of some new pyrazole incorporated imidazole derivatives. **European Journal of Medicinal Chemistry**. 2011; 46: 3531-3536.

14. Yang X, Wan W, Deng X, Li Y, Yang L, Li L and H. Zhang H. Design, synthesis and cytotoxic activities of novel hybrid compounds between 2-phenylbenzofuran and imidazole. **Bioorganic and Medicinal Chemistry Letters**. 2012; 22: 2726-2729.
15. Lu X, Liu X, Wan B, Franzblau SG, Chen L, Zhou C and You Q. Synthesis and evaluation of anti-tubercular and antibacterial activities of new 4-(2,6-dichlorobenzyloxy)phenyl thiazole, oxazole and imidazole derivatives. **European Journal of Medicinal Chemistry**. 2012; 49: 164-171.
16. Zhan P, Liu X, Zhu J, Fang Z, Li Z, Pannecouque C and Clercq ED. Synthesis and biological evaluation of imidazole thioacetanilides as novel non-nucleoside HIV-1 reverse transcriptase inhibitors. **Bioorganic and Medicinal Chemistry**. 2009; 17: 5775-5781.
17. Nakagawa T, Yokozawa T, Terasawa K, Shu S and Juneja LR. Protective activity of green tea against free radical- and glucose-mediated protein damage. **Journal of Agricultural Food Chemistry**. 2002; 50: 2418-2422.
18. Weatherburn MW. Phenol-hypochlorite reaction for determination of ammonia. **Analytical Chemistry**. 1967; 39: 971-974.
19. Rakesh KP, Shantharam CS and Manukumar HM. Synthesis and SAR studies of potent H⁺/K⁺-ATPase inhibitors of quinazolinone-schiff's base analogues. **Bioorganic Chemistry**. 2016; 68: 1-8.
20. Rakesh KP, Manukumar HM and Gowda DC. Schiff's bases of quinazolinone derivatives: synthesis and SAR studies of a novel series of potential anti-inflammatory and antioxidants. **Bioorganic and Medicinal Chemistry Letters**. 2015; 25: 1072-1077.